Characterization and Antimicrobial Activity of Silver Doped Hydroxyapatite Obtained by the Microwave Method

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corres@http://dx.doi.org/10.5755/j01.ms.22.3.12133

Received 27 April 2015; accepted 20 August 2015

In this study, silver (Ag) doped hydroxyapatite (HA) was produced by the microwave method and its antimicrobial activity was investigated. The physical, chemical and structural characteristics of the material were determined using multi-purpose X-ray diffractometry (XRD), Fourier transform infrared spectrometry (FTIR), and scanning electron microscope-energy dispersive X-ray spectroscopy (SEM-EDS) apparatus. The amount of silver in the solutions of silver-doped hydroxyapatite obtained were determined with the use of an inductively coupled plasma optical emission spectroscopy instrument (ICP-OES). The minimum inhibition concentration (MIC) of the silver doped hydroxyapatite (Ag-HA) against the test microorganisms was determined by the Broth Microdilution method. It was established that a concentration of 2.09 – 12.25 µg/ml was effective against gram-negative bacteria (Escherichia coli ATCC 12228, Salmonella typhimurium CCM 5445, Pseudomonas aeruginosa ATCC 27853, Klebsiella pneumoniae CCM 2318), and 4.18 – 12.25 µg/ml was effective against gram-positive bacteria (Staphylococcus aureus ATCC6538-P, Bacillus subtilis ATCC 6633, Enterococcus faecalis ATCC 29212) and the yeast Candida albicans ATCC 10239.

Keywords: silver, hydroxyapatite, microwave, MIC value, antimicrobial activity.

1. INTRODUCTION

The importance of antimicrobial ceramics, which are used widely in factories, building materials, cosmetics and electrical appliances, is steadily increasing. These materials need to have a supporting structure to which metal ions can easily be added. The basal material can be classified as amorphous silica, zeolite or calcium phosphate, which shares the characteristic of a broad crystal structure. Thanks to this, the metal ions enter the system and affect microorganisms [1, 2].

Calcium phosphate, known as hydroxyapatite (HA) has the structural formula Ca10(PO4)6(OH)2. Several chemical and thermal methods have been developed to produce this compound in the form of ceramic powder [3, 4, 5]. Because of the calcium and phosphorus which it contains, HA has a high level of compatibility with the human body, and as a result is used in various parts of the body as an implant. At the same time, the exchange rate of metal cations such as Ag+, Cu2+, Zn2+ in HA is very rapid [6].

It has long been known that silver and silver ions have a strong antimicrobial effect. Because of this effect and because they are non-toxic, silver ions are applied as an additive to coating materials to the surfaces of many items such as ceramics, glass, tilework, plastic, paper, paint or textiles which are in daily use, but have many microorganisms, either at the time of manufacture or later [2].

Silver doped HA (Ag-HA) can be obtained by various methods such as chemical precipitation, sol-gel and ion exchange [7, 8, 9]. However, in the present study it was obtained by a different method using microwaves, and the antimicrobial effect of the Ag-HA obtained was tested by the Broth Microdilution method against eight microorganisms [10].

2. MATERIALS AND METHOD

2.1. Preparation and characterization of Ag-HA solutions

In the preparation of Ag-HA solutions, calcium hydroxide (Ca(OH)2), diammonium hydrogen phosphate ([NH4]2HPO4) and silver nitrate (AgNO3) were used as precursor materials. Ultra pure water was used as a solvent in the experiments.

Solutions of pure HA and Ag-HA were prepared by the microwave method according to [11]. AgNO3 (0.25 M) was dissolved in pure water at room temperature. Then a 0.3 M suspension of calcium hydroxide was prepared in a different glass flask by mixing with pure water. Following this, the silver nitrate solution was added dropwise to the calcium hydroxide suspension and 0.3 M diammonium phosphate was added to this solution, with five minutes of strong stirring. The prepared solution was then exposed to 30 minutes of heat treatment in a microwave oven (800 W, 2.5 GHz). In this way, colloidal hydroxyapatite solutions containing various concentrations of silver (0.5 – 5 %) were obtained, and were used in the subsequent tests. In order to perform characterization analyses, the Ag-HA solutions were dried at 100 °C for 12 hours in a drying oven and ground to a powder in a mortar. Pure hydroxyapatite, which was used as a control, was prepared by the same method but without the addition of silver nitrate.
The physical, chemical and structural characteristics of the materials obtained were determined by means of X-ray diffractometry (XRD, Philips), Fourier transform infrared spectrometry (FTIR, Perkin Elmer) and Scanning electron microscope-Energy dispersive X-ray spectroscopy instruments (SEM-EDS). For phase analysis, XRD of samples was carried out under Cu-Kα radiation and data were taken at a rate of 2°/min scanned at 2θ = 10°–70° interval. The structure of the HA and Ag-HA powders was determined by FTIR spectrum scanning in the range of 4000–400 cm⁻¹. Scanning electron microscope (JEOL JSM-6060) was used to determine the surface morphology of the hydroxyapatite powders. The amount of silver in the hydroxyapatite solutions obtained was determined by means of an inductively coupled plasma optical emission spectroscopy (ICP-OES, Perkin Elmer) instrument.

2.2. Determination of MIC values of hydroxyapatite solutions

In order to establish the in vitro antimicrobial activity of the 0–5% solutions of Ag-HA prepared, the Gram-positive bacteria Escherichia coli ATCC 12228, Salmonella typhimurium CCM 5445, Pseudomonas aeruginosa ATCC 27853, Klebsiella pneumoniae CCM 2318; the Gram-negative bacteria Staphylococcus aureus ATCC6538-P, Bacillus subtilis ATCC 6633, Enterococcus faecalis ATCC 29212 and the yeast Candida albicans ATCC 10239 were used. The minimum inhibition concentrations (MIC) of the solutions were determined with 96-well microplates using the Broth Microdilution technique [10]. Activation of microorganisms was obtained by incubation in Müller Hinton Broth (MHB) at 37 °C for 18–20 hours [12, 13, 14].

The 0–5% Ag-HA solutions were tested in a doubly diluted form. A certain aliquot of each dilution and MHB containing each test microorganism were poured into each well to make the wells up to 200 μL. After 24 hours of incubation, 50 μL of 0.5% TTC (triphenyl tetrazolium chloride) was added on top of the cells, and incubation was continued for another two hours. At the end of this period, the red formazan compound formed by the reduction of the TTC was accepted as an indicator of reproduction, and used to determine the MIC value [15]. The MIC was determined as the lowest concentration that inhibited the visible growth of the test microorganism.

3. RESULTS AND DISCUSSION

3.1. X-ray diffraction phase

Fig. 1 shows the XRD diffraction patterns for the pure HA and Ag-HA powders. The peaks in these graphs were compared with data in the JCPDS catalogue [16] and they showed characteristic peaks which were consistent with the peaks given for hydroxyapatite by the International Diffraction Data Centre. Here, the characteristic peaks of the solid phases of HA formed by the pure HA ceramic (2θ ~ 25–34.5°) clearly showed the development of a weak crystallized apatite phase. It was clearly seen that all the peaks relating to hydroxyapatite in the powder samples in Fig. 1 at 20 were generally at 25.80°, 28.60°, and 32.10°. In addition, at 20, low density peaks were found at 34.20°, 39.60°, 46.50° and 49.00°. In the XRD results of HA samples with 5% Ag, the characteristic peaks of silver oxide (Ag₂O) were observed along with those of hydroxyapatite.

Chung et al. observed that the phase characteristic of HA formed when it calcined at a temperature of 350 °C, and that Ag₂O and CaO peaks appeared when the amount of Ag was 2000 ppm [8]. These studies used silver nitrate as a source of silver, and they used the sol-gel method at a low temperature, so that it was shown that the formation of Ag₂O arose from electrostatic attraction to HA. In a study by Rameshbabu et al., antibacterial Ag-HA was synthesized by the microwave method, and it was determined from the XRD results that all patterns corresponded to the hexagonal crystals of HA in the JCPDS 9-432 database, and were almost the same [11]. However, they reported that in samples where the amount of silver was over x ≥ 0.4, the 2θ ~ 36.68° (Ag₃PO₄, JCPDS 6-505) silver phosphate peaks were not clearly visible. The 0–5% Ag-HA obtained in the present study was not subjected to any heat treatment other than drying in an oven at 100 °C. As the amount of silver attaching to the HA under these conditions was 5%, no impurity phase was encountered apart from observation of the Ag₂O phase.

3.2. Scanning electron microscope (SEM) analysis

Fig. 2 shows the results of SEM analysis of the synthesized pure HA and Ag-HA.

Fig. 1. XRD patterns of the 0%–5% Ag-HA

Fig. 2. SEM micrographs of HA (a) and 1.5% Ag-HA (b)

In images taken by the secondary electron imaging technique of the scanning electron microscope, powder containing pure HA had fine and rounded lines, while Ag-HA powder showed morphologically more distinct lines. The results of SEM analysis showed that when the microstructure obtained was analyzed the powders generally had a particle size of 50 μm or less (Fig. 2). It was seen that the synthesized HA powder did not show a
homogeneous distribution because it had been ground manually in a mortar. It was determined that as the size of the particles in the powders in the study decreased, they tended to come together and form agglomerates in order to reduce the increasing surface energies.

Authors found that the average particle size in Ag-HA which they obtained by the sol-gel method was between 3 and 19 µm [8]. Others stated that particles of pure HA and 0.5 % Ag-HA synthesized with the use of microwaves were needle-shaped and showed a great tendency to flocculate [10]. However, they reported that 5 % Ag-HA particles formed more regular lines and heaped up less, which could be an effect of the silver phosphate phase around them. The SEM results obtained in the present study show that particles of Ag-HA produced by the microwave method were small in dimensions and had a homogeneous distribution.

3.3. Energy dispersive X-ray spectroscopy analysis

The effective antimicrobial agent trapped in the crystal structure of the powder, that is the silver cations, were determined using Energy dispersive X-ray spectroscopy (EDS). Results are shown in Fig. 3. The values obtained by EDS were compared with stoichiometric values measured during powder production, and it was found that the Ca/P ratio in the sample of pure HA (1.667) was close to the stoichiometric HA ratio (Ca/P = 1.67), and that this ratio was 1.573 in the sample of 1 % Ag-HA (Fig. 3). In addition, elemental analysis ascertained the presence of Ag in Ag-HA.

![Fig. 3. SEM/EDS spectra of HA (a) and 1.0 % Ag-HA (b)](image)

Feng et al. examined the morphology and composition of Ag-HA coating by SEM and EDS, and found that the Ca/P atomic ratios of silver-doped and non-silver-doped coatings were 1.19 and 1.46 respectively [17]. The researchers concluded that the Ca²⁺ ions in the HA structure had been exchanged for Ag⁺ ions. In the present study, it was thought that the difference found in the Ca/P ratios in the 1 % Ag-Ha sample may stem from the exchange of places of the Ca ions in the HA cage with Ag ions, and losses during the heat treatment at the time of synthesis of the powders.

3.4. ICP-OES analysis

A determination of the silver content of Ag-HA was performed by ICP-OES analysis, and the values for pure HA and Ag-HA are given in Fig. 4. According to these results, the ICP values are in proportion to the previously calculated amounts of silver (pure HA, 0.5 % Ag-HA and 1 % Ag-HA) and as the amount of silver increased, so this ratio increased in proportion (2 % Ag-Ha and 3 % Ag-HA). It was observed that the amounts of Ag doped into the hydroxyapatite structure also showed differences according to the HA ion exchange capacity (5 % Ag-HA).

![Fig. 4. Amounts of Ag in HA samples](image)

Oh et al. doped Ag ions into HA powder by immersion in a solution of AgNO₃, and reported that although a slight increase in the amount of doped silver was observed with the increase in the arranged amount of silver, after drying by evaporation 2.21 % of Ag, and after freeze drying 4.87 % of Ag, were doped, the opposite to the assumption that the HA would ion-exchanged with 15 % of the Ag out of the total moles of cations [7]. In another study, the amount of Ag in Ag-HA obtained by the precipitation method was determined using an ICP apparatus, and an increase in the amount of doping Ag was observed with an increase in the Ag/Ca ratio [9]. It was found that the Ag concentration in the HA was not in accord with the concentrations in the starting materials; however, the values were within acceptable deviation ratios, and most importantly, increases were found [8]. In our study, there was an increase in the amounts of silver calculated for Ag-HA and doped Ag, and it is thought that this came about as a result of ion exchange between Ag ions and Ca ions in the HA cage causing a reduction in Ca ions and an increase in Ag ions. However, when the amount of Ag in the HA was 5 %, a reduction in the amount of doped Ag indicated that the amount of exchangeable Ca²⁺ ions in the hydration layer of the HA was only about 4 % of the total Ca²⁺ ions in the crystals.
3.5. Findings of FTIR analysis

Table 1 shows spectra and observed functional groups which characterize the bond structures of HA and silver-doped HA powders, as obtained by FTIR. FTIR spectra showed that the $v_1$ PO$_4^{3-}$ band around 962 cm$^{-1}$, the $v_2$ PO$_4^{3-}$ band around 472 cm$^{-1}$, the $v_3$ PO$_4^{3-}$ band around 1025 and 1090 cm$^{-1}$ and the $v_4$ PO$_4^{3-}$ band around 562, 559 and 601 cm$^{-1}$ are the important characteristic bands of hydroxyapatite, and the sharp peaks at 3570 and 632 cm$^{-1}$ were the O-H band (Table 1).

It was determined that bands at ~1090 and 962 cm$^{-1}$ reflected the P-O vibration modes indicating a weakly crystallized hydroxyapatite structure, and that the band at ~602 cm$^{-1}$ reflected the hydroxyapatite peaks which correspond to the tension modes of (PO$_4^{3-}$) ions [18]. Su and Xiong examined the FTIR spectra of HA heat-treated at 600 °C and Ag-HA which had not undergone heat treatment, and reported that the sharp peaks at 3571 and 631 cm$^{-1}$ were related to O-H vibration, and that the peaks at 1090, 1041, 962, 601, 567 and 474 cm$^{-1}$ reflected PO$_4^{3-}$ vibration [19]. Rameshbabu et al. observed the characteristic peaks for HA in the spectra of Ag-HA dried in an oven and heat-treated at 900 °C in two samples, PO$_4^{3-}$ tension and bending modes at 900–1200 cm$^{-1}$, PO$_4^{3-}$ bending mode at 602 cm$^{-1}$, and O-H vibration mode at 632 and 3571 cm$^{-1}$ [11].

Table 1. The functional groups in FTIR spectra of the 0%–5% Ag-HA in as synthesized condition

<table>
<thead>
<tr>
<th>Functional groups</th>
<th>HA</th>
<th>0.5% Ag-HA</th>
<th>1% Ag-HA</th>
<th>1.5% Ag-HA</th>
<th>2% Ag-HA</th>
<th>3% Ag-HA</th>
<th>5% Ag-HA</th>
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<tr>
<td>$v_2$ PO$_4^{3-}$ vibration band</td>
<td>471</td>
<td>471</td>
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<td>471</td>
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<tr>
<td>$v_3$ PO$_4^{3-}$ vibration band</td>
<td>562</td>
<td>563</td>
<td>560</td>
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<tr>
<td>$v_4$ PO$_4^{3-}$ vibration band</td>
<td>600</td>
<td>602</td>
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<tr>
<td>$v_1$ PO$_4^{3-}$ vibration band</td>
<td>962</td>
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<tr>
<td>OH band</td>
<td>1025</td>
<td>1025</td>
<td>1025</td>
<td>1024</td>
<td>1024</td>
<td>1024</td>
<td>1023</td>
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<tr>
<td>OH band</td>
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<td>632</td>
<td>632</td>
<td>633</td>
<td>633</td>
<td>631</td>
<td>632</td>
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<tr>
<td>OH band</td>
<td>3570</td>
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</table>

*not shown in Fig. 5.*

Shirkhanzadeh at al. stated that no extra peaks were observed in the FTIR spectrum between the wavelengths of 400 and 4000 cm$^{-1}$ when HA was doped with Ag [20]. Nath et al. stated that the Ag-O band of Ag-O and AgO phases could not be detected by the 400–4000 cm$^{-1}$ wavelength FTIR scanning limit and therefore apart from the O-H vibration peak change at 3570 cm$^{-1}$, FTIR results did not show the presence of silver ions [21]. Thus, it was shown that the O-H group in the HA cage is modified by the presence of an Ag ion. In our study, the characteristic peaks for hydroxyapatite in the FTIR spectra of both pure HA and Ag-HA were in accordance with the literature. However, the 3570 cm$^{-1}$ peak for the hydroxyl group in the FTIR spectrum of the compound Ag-HA was not observed (Fig. 5). It is thought that the reason for this is that as explained above, the silver substitutions in the HA take place in accordance with the formula $Ca_xAg_y(PO_4)_3(OH)_2$.

3.6. Results of MIC values of 0–5% Ag-HA solutions

The calculated MIC values of solutions of pure HA and 0.5–5% Ag-HA against gram-negative and gram-positive bacteria and C. albicans are given in Table 2. According to the results obtained, pure HA and 0.5% Ag-HA showed no antimicrobial activity to any of the test organisms, while solutions of 1–5% Ag-HA showed different activities according to the species of microorganism. Effective MIC values against gram-negative bacteria were 2.09–12.25 μg/ml, and against gram-positive bacteria and C. albicans they were 4.18–12.25 μg/ml. MIC concentrations effective against the gram-negative bacterium K. pneumoniae and P. aeruginosa were recorded respectively as 2.09–5.38 and 2.09–9.10 μg/ml.

Table 2. MIC values of 0–5% solutions of Ag-HA against bacteria and C. albicans (μg/mL)

| Organism | MIC (μg/mL) | Ag-HA 0.5% | | Ag-HA 1% | | Ag-HA 1.5% | | Ag-HA 2% | | Ag-HA 3% | | Ag-HA 5% |
|----------|------------|------------|----------|-----------|----------|-----------|----------|-----------|----------|-----------|----------|
| E. coli  | –          | 10.76      | 9.10     | 6.12      | 8.37     | 11.50     |
| S. typhimurium | –       | 10.76 | 9.10 | 12.25 | 8.37 | 11.50 |
| P. aeruginosa | –        | 5.38 | 4.55 | 6.12 | 2.09 | 2.87 |
| K. pneumoniae | –         | 5.38 | 9.10 | 3.06 | 2.09 | 2.87 |
| S. aureus | –          | 10.76 | 9.10 | 6.12 | 4.18 | 5.75 |
| S. epidermidis | –        | 10.76 | 9.10 | 12.25 | 8.37 | 5.75 |
| E. faecalis | –         | 10.76 | 9.10 | 6.12 | 4.18 | 5.75 |
| C. albicans | –         | 10.76 | 9.10 | 6.12 | 4.18 | 5.75 |

Feng et al. examined the antibacterial effect of a thin film of Ag-HA on an aluminium substrate against gram-negative E. coli and P. aeruginosa and gram-positive S. aureus and S. epidermidis, and it was determined that a 12.7 ppm concentration of Ag in HA prevented reproduction of the test bacteria on the aluminium substrate, and killed the bacteria [17].

Oh et al. compared the MIC values of Ag-HA obtained by the ion-exchange and precipitation methods against the
gram-positive *S. aureus* and the gram-negative *E. coli* bacteria, and determined that as the amount of silver increased, the MIC values of Ag-HA prepared by both methods against both bacteria was effective at higher concentrations [9]. However, differences were observed in the MIC values against both bacteria between Ag-HA with approximately the same amount of silver according to the synthesis method. Nirmala et al. stated that the bactericidal effects of Ag-HA were more pronounced with an increased amount of Ag, and that HA containing 1% of silver showed an MIC value against *S. aureus* varieties which were either resistant or susceptible to methicillin, while HA containing 3% silver showed an MIC value against *E. coli* [22].

Two mechanisms have been developed to explain the antimicrobial effects of Ag-HA. First, that microorganisms adhere to the surface by electrostatic force and interact directly with metal ions. The second mechanism is that the slow release of Ag from HA can inhibit bactericidal activity in the medium [23–25].

Several studies have been carried out to demonstrate the antimicrobial activity of Ag-HA powders as well as the surfaces of Ag-HA coatings against a broad range of microorganisms, especially bacteria, and different results have been obtained [23, 26, 27, 28]. The broad antibacterial spectrum of Ag-HA is similar to the spectrum of silver ions, and like silver ions, it has a more effective influence on gram-negative rather than gram-positive bacteria [26]. The reason for different sensitivities between gram-positive and gram-negative bacteria and *C. albicans* can be found in the different transparencies of the cell wall [29]. The cell wall of gram-positive bacteria consists of peptidoglycans (mureins) and teichoic acids, while the cell wall of gram-negative bacteria consists of lipo polysaccharides and lipopolysaccharides, and that of fungi consists of polysaccharides such as hitchin and glucan [30, 31].

Apart from this, the antimicrobial effects of Ag-HA may be associated with the characteristics of specific bacterial strains. Thus, Bai et al. (2012) reported that a higher content of Ag in functionally graded HA coatings particularly 3% and 6.5 wt.% caused a reduction in the viability of *S. aureus* (Cowan I strain, ATCC 12598), as opposed to 1 wt.% of Ag [27]. Chen et al. (2007) evaluated the surface of Ag-HA coatings against *S. aureus* (Cowan I) and observed the reduced adhesion of *S. aureus* on 1% Ag-HA when compared to a 1.5% Ag-HA coating [26]. Samani et al. (2013) determined that 1.5% of Ag in an HA coating has a strong antibacterial effect against *S. aureus* (Methicillin-resistant) due to the powerful antibacterial property of Ag [23]. Also, Colbanu et al. (2011) revealed lower adhesion of *S. aureus* (ATCC 6538) on nano-sized xAg-HA powder with x = 0.2 content of Ag [28].

The reported results are higher than those obtained by us in the present study, which suggests that the antimicrobial activity of Ag-HA may be influenced by the bacterial strains used as well as by the preparation method. The method of preparation or the possible treatments results in apatites, which have a characteristic degree of crystallinity and a specific surface area. It is well known that the antibacterial activity of silver ions increases with surface area, due to its extent of release from the apatite. Therefore, the methods by which the fine crystals are obtained lead to obtaining a material displaying more powerful antibacterial activity [25]. Thus, the relationship between the method of synthesis of the Ag-HA and its antimicrobial effect should also be examined.

4. CONCLUSIONS

In this study, an investigation was made of the antimicrobial effectiveness of silver-doped hydroxyapatite synthesized by the microwave method. The physical, chemical and structural characteristics of the material were determined using multi-purpose X-ray diffractometry (XRD), Fourier transform infrared spectrometry (FTIR), and scanning electron microscope-energy dispersive X-ray spectroscopy (SEM-EDS) apparatus. The amount of silver in the solutions of silver-doped hydroxyapatite obtained were determined with the use of an inductively coupled plasma optical emission spectroscopy instrument (ICP-OES), and it was established that the amount of added silver and the calculated amount were proportional. The effective concentrations of the silver-doped hydroxyapatite against test microorganisms was determined by means of the broth microdilution technique, and the effective MIC value against gram-negative bacteria (*E. coli, S. thiphimurium, P. aeruginosa* and *K. pneumoniae*) was found to be 2.09–12.25 μg/ml, and against gram-positive bacteria (*S. aureus, B. subtilis, and E. faecalis*) and *C. albicans* it was 4.18–12.25 μg/ml. In the light of these results, future studies will concern Ag-HA as an antibacterial coating additive on leather.

Acknowledgement

We would like to thank the Research Foundation of Ege University for their financial support (Project No 09MÜH080).

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